**Taxonomic status of Atocion hypanicum (Klokov) Tzvelev (Caryophyllaceae) inferred from analysis of ITS1 and ITS2 secondary structure**

**VIKTORIJA О. MARTynyuk, OKsANA V. TYSCHchenko, NATALIJA I. KARPENKO, ANDRII S. TARIEiev, IHEOR Yu. KOSTIKOV**


Secondary structures of ITS1 and ITS2 transcripts of Atocion hypanicum (Klokov) Tzvelev and A. compactum (Fisch. ex Hornem.) Tzvelev were reconstructed. The presence of a unique nucleotide substitution in fourth helix of A. hypanicum ITS1 transcript, which lead to a disturbance in the secondary structure of the helix, strongly supported it as a separate operational taxonomic unit. The lack of CBC in obtained sequences of ITS2 from A. hypanicum and A. compactum confirmed the absence of genetic barriers for the taxa crossing. Phenotypical distinctions between A. hypanicum and A. compactum from Caucasus associated with bracts and the morphology of the uppermost leaves are demonstrated. The separate species status of A. hypanicum was confirmed.

**Key words:** Atocion hypanicum, A. compactum, ITS1 and ITS2 secondary structure, taxonomic status


Проведено реконструкцію вторинних структур послідовностей ITS1 та ITS2 транскриптів Atocion hypanicum (Klokov) Tzvelev та A. compactum (Fisch. ex Hornem.) Tzvelev. Наявність унікальної нуклеотидної заміни у четвертій спіралі транскрипту послідовності ITS1 A. hypanicum, що призводить до порушення вторинної структури спіралі, вказує на статус цього таксону як окремої операційної таксономічної одиниці. Відсутність CBC в послідовності ITS2 між отриманими сіквентами A. hypanicum та A. compactum свідчить про відсутність генетичних бар’єрів для схрещування цих таксонів. Показано наявність фенотипічних відмін між A. hypanicum та A. compactum з території Кавказу, пов’язаних з морфологією приквіток та верхівкових листків. Підтверджено самостійний видовий статус A. hypanicum.

Ключові слова: Atocion hypanicum, Atocion compactum, вторинна структура ITS1 та ITS2, таксономічний статус

Taxonomic status of *Atocion hypanicum* (Klokov) Tzvelev (Caryophyllaceae) inferred from analysis of ITS1 and ITS2 secondary structure

*Atocion hypanicum* (Klokov) Tzvelev (basionym *Silene hypanicca* Klokov) was described in 1948 by M.V. Klokov from Pobuzhya granite outcrops as a new species *S. hypanicca* inside the genus *Silene* L. [KLOKOV, 1948]. Klokov also noted that earlier on the territory of Ukraine this plant was identified as *S. compacta* Fisch. – a species, described by F. Fischer in 1812 from Caucasus. Additionally, he noticed that *S. hypanicca* is an endemic of granitic outcrops of Pivdenny Bug. In 2001 *S. hypanicca* was transferred to the genus *Atocion* by N.N. Tzvelev [TZELEV, 2001]. In agreement with his predecessors, [FLORA..., 1952; MOSYAKIN, FEDORONCHUK, 1999], he considered *A. hypanicca* to be a separate species, related to *A. compacta* (Fisch. ex Hornem.) Tzvelev (basionym *Silene compacta*).

In the domestic literature *Atocion hypanicca* is now considered to be a strict local endemic of Pivdenny Bug, included in different Ukrainian issues of nature conservation and referred as “vulnerable” [OPREDELTITEL...., 1987; CHERVONA..., 1996, 2009; FEDORONCHUK, DIDUKH, 2002]. Several localities of this species are known. All of them are located on the south spurs of Pryniprovska highland, within the boundaries of Right Bank Steppe, in Pivdenny Bug valley. Local populations have small area, compact-diffuse structure and are stably decreasing [FEDORONCHUK M.M., DIDUKH YA. P. et al., 2002; CHERVONA..., 2009]. In 1991 *Silene hypanicca* was placed to European Red List [EUROPEAN ..., 1991], but in 2011 [BILZ et al, 2011] this species was not included there.

Likely, the reason for *S. hypanicca’s* removal from the European Red List is change in the understanding of *S. compacta* volume in 1990-s. In particular, *S. hypanicca* was considered as a probable synonym of *S. compacta* in West-European literature [TUTIN et al., 1993]. It should be mentioned, that the tendency for the enlargement of *S. compacta* appeared earlier. In 1967 *S. vandasii* Nábělek, described in 1923 from south-east Turkey [FLORA ..., 1967], was included into *S. compacta*. Additionally, we emphasize, that both viewpoints: for separateness of *S. hypanicca* and volume of *S. compacta* were based exclusively on the results of comparison of morphological descriptions and herbarium specimens.

In 2009, in order to unravel the phylogenetic relationships between genera *Atocion* and *Viscaria* Bernh., a molecular analysis was performed. 6 sequences of nuclear (including ITS1 and ITS2) and chloroplast DNA markers were obtained for five samples, named *Atocion compactum* by the authors [FRAJMAN et al., 2009]. The material, used in this research, was collected on the territory of Turkey, Georgia and Macedonia. Moreover, one sample, described as cultivar seeds of *A. compactum* from Ukraine¹, but without any notices about its possible connection to *A. hypanicca* was investigated in the study. The material from *A. hypanicca* type locality was not yet included in the research. Therefore, the questions

¹ According to NCBI annotation this material is received from UPS herbarium (specimen 330 from A.Rautenberg collection) which according to publication is called “Cultivated: seeds from Ukraine” [FRAJMAN et al., 2009]. However, according to collector’s private label specimen source is unknown (www.sileneae.info). Further we will call this specimen “Ukrainian cultivar”.

Ключевые слова: *Atocion hypanicum*, *Atocion compactum*, вторичная структура ITS1 и ITS2, таксономический статус
about status and volume of *A. compactum*, as well as separateness of *A. hypanicum* were left without discussion. Hence, the taxonomical status of *A. hypanicum* remains unclear.

The purpose of our research was to clarify the taxonomical status of *A. hypanicum* on the basis of comparative analysis of the original molecular data obtained from type locality of *A. hypanicum* on the territory of Ukraine and *A. compactum* from Caucasus, and also sequences from the same species deposited in NCBI. We analyzed the structure of marker sequences from ITS1-5.8S-ITS2 cluster of ribosomal genes with estimation of taxonomic weights of discriminant phenotypic traits between *A. compactum* and *A. hypanicum*.

**Matherials and methods**

Material: the original samples from the type locality of *A. hypanicum* (vicinity of Bogdanivka, Domanivka distr., Mykolaiv reg., Ukraine) and *A. compactum* from Caucasus (vicinity of Dombay, Karachay–Cherkess Republic, Russian Federation), and also 29 herbarium specimens from two herbaria of Ukraine (KW and KWU).

Leaves' fragments of herbarium specimen of *A. hypanicum* (KW №081628) and seedlings of *A. compactum* were used for molecular analysis. Total DNA extraction was performed using a modified CTAB-method [TARIEIEV et al., 2011]. ITS1-5.8S-ITS2 sequence amplification of *A. hypanicum* was performed using ITS1 and ITS4 primers. The same sequence of *A. compactum* was obtained using ITS4 and ITS5 primers (table 1) on Techne thermocycler according to White [WHITE, 1990]. Sequencing of amplicons with forward and reverse primers was done commercially in Macrogen Inc. (http://www.Macrogen.com, Netherlands).

<table>
<thead>
<tr>
<th>Назва праймера</th>
<th>Послідовність</th>
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<tbody>
<tr>
<td>ITS1-forward</td>
<td>5'-TCCGTAGGTGAACCTGCGG-3'</td>
</tr>
<tr>
<td>ITS4-reverse</td>
<td>5'-TCCCTCGCTTATGTATGC-3'</td>
</tr>
<tr>
<td>ITS5-forward</td>
<td>5'-GGAAGTAAAAGTCCGTAACAGG-3'</td>
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</tbody>
</table>

Sequences were edited with BioEdit software [HALL, 1999] through forward and reverse chromatogram comparison. Obtained sequences were deposited in NCBI (accession number KJ616753 and KM975935).

ITS1 sequence was annotated in accordance to *A. compactum* (accession number FJ384030), ITS2 annotation was done by the modeling of terminal part of 5.8S sequence and its complementary part of 28S in MFold [ZUKER, 2003]. Screening of NCBI nucleotide collection was performed using BLAST (blast.ncbi.nlm.nih.gov) [ALTSCHUL et al., 1990]. The data set for comparison of secondary structures included original data on *A. compactum* and *A. hypanicum* as well as other sequences of *A. compactum* from GeneBank with the next NCBI accession numbers: FJ384028 and FJ384029 (Turkey), FJ384030 (Georgia), FJ384031 (Macedonia), FJ384032 (“Ukrainian cultivar”). ITS2 secondary structure reconstruction was done by online service MFold [ZUKER, 2003]. Obtained structures were visualized in Pseudoviewer 3.0 [BYUN et al., 2009].

Comparative morphological analysis was performed on the basis of originally collected material and herbarium samples mentioned above. The sample of cultivar of *A. compactum*, collected in Sevastopol (KW №014166) was also included in the research.

**Results**

**Molecular-genetic analysis.** Obtained ITS1-5.8S-ITS2 sequences of *A. hypanicum* (646 bp.) and *A. compactum* (628 bp.) were most similar (over 98 %) to five sequences, which
were deposited as *A. compactum* (NCBI accession numbers FJ384028, FJ384029, FJ384030, FJ384031, FJ384032), according to the results of BLAST search. The sequences of *A. compactum* from the territory of Macedonia (FJ384031) and Turkey (FJ384029) were most similar to *A. hypanicum* (99.69 % and 99.36 % of identity, respectively). The sequences of samples from Georgia (FJ384030) and so called “Ukrainian cultivar” (FJ384032) displayed lower similarity (98.76 % and 99.07 %, respectively). The last sample (FJ384028, Turkey) shared 98.89 % of similarity, but all eight different positions contained ambiguously identified nucleotides. The similarity of original *A. compactum* sequence to the described sequences was as following: FJ384031 – 100 %, FJ384032 – 99,36 %, FJ384030 – 99,04 %, FJ384029 – 99.68 % and FJ384028 – 98.88 %.

In total 10 variable positions for ITS1 and 4 – for ITS2 were detected in sampling (tab. 2).

**Table 2**  
Variable sites in ITS1 and ITS2 sequences of *A. hypanicum* and *A. compactum*  

<table>
<thead>
<tr>
<th>Position of variable site in sequence</th>
<th>58</th>
<th>69</th>
<th>74</th>
<th>77</th>
<th>80</th>
<th>102</th>
<th>125</th>
<th>133</th>
<th>210</th>
<th>216</th>
<th>40</th>
<th>80</th>
<th>130</th>
<th>189</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>A. hypanicum</em></td>
<td>C</td>
<td>T</td>
<td>C</td>
<td>C</td>
<td>G</td>
<td>C</td>
<td>C</td>
<td>S (C)</td>
<td>G</td>
<td>C</td>
<td>A</td>
<td>C</td>
<td>T</td>
<td></td>
</tr>
<tr>
<td>FJ384031 (UA)</td>
<td>C</td>
<td>T</td>
<td>C</td>
<td>C</td>
<td>G</td>
<td>C</td>
<td>C</td>
<td>G</td>
<td>G</td>
<td>C</td>
<td>A</td>
<td>C</td>
<td>T</td>
<td></td>
</tr>
<tr>
<td><em>A. compactum</em> (RU)</td>
<td>C</td>
<td>T</td>
<td>C</td>
<td>C</td>
<td>G</td>
<td>C</td>
<td>C</td>
<td>G</td>
<td>G</td>
<td>C</td>
<td>A</td>
<td>C</td>
<td>T</td>
<td></td>
</tr>
<tr>
<td>FJ384030 (GE)</td>
<td>C</td>
<td>T</td>
<td>C</td>
<td>C</td>
<td>G</td>
<td>C</td>
<td>C</td>
<td>G</td>
<td>G</td>
<td>C</td>
<td>A</td>
<td>C</td>
<td>T</td>
<td></td>
</tr>
<tr>
<td><em>A. hypanicum</em></td>
<td>Y</td>
<td>T</td>
<td>C</td>
<td>Y</td>
<td>G</td>
<td>Y</td>
<td>M</td>
<td>G</td>
<td>Y</td>
<td>W</td>
<td>C</td>
<td>T</td>
<td></td>
<td></td>
</tr>
<tr>
<td>FJ384032 (TBC)</td>
<td>C</td>
<td>C</td>
<td>C</td>
<td>C</td>
<td>T</td>
<td>G</td>
<td>C</td>
<td>G</td>
<td>G</td>
<td>C</td>
<td>A</td>
<td>T</td>
<td>C</td>
<td></td>
</tr>
<tr>
<td><em>A. hypanicum</em></td>
<td>SNP</td>
<td>nst,S</td>
<td>NP</td>
<td>hCBC</td>
<td>SNP</td>
<td>hCBC</td>
<td>SNP</td>
<td>SNP</td>
<td>SNP</td>
<td>SNP</td>
<td>sst</td>
<td>sst</td>
<td>SNP</td>
<td>nst</td>
</tr>
<tr>
<td>Abbreviations: SNP – single nucleotide polymorphism; nst – nonstructural substitution; sst – structural substitution; hCBC – hemicompensatory base change.</td>
<td></td>
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</tbody>
</table>

The results of ITS1 secondary structure comparison of *A. hypanicum* and *A. compactum* and also *A. compactum* sequences, deposited in NCBI, revealed that analyzed sequences are heterogeneous. Two hemicompensatory base changes are present in positions 74 and 80 of FJ384029 and FJ384032 respectively. Also one nonstructural substitution in the terminal part of the first helix of *A. compactum* (FJ384032, position № 69) was observed (fig. 1) and a range of ambiguous sites (positions № 58, 69, 77, 80, 125 and 133) were detected in some sequences of *A. compactum*.

*A. hypanicum* can be distinguished from all analyzed *A. compactum* sequences by the substitution in position № 210 in ITS1 sequence. In this position cytosine was present for *A. hypanicum* (instead of guanine for *A. compactum*). The presence of guanine minor peak in the sequence of *A. hypanicum* (fig. 2) can be argued by polycopycity of ribosomal genes. Nucleotide substitution in position № 210. C>G was structural (sst) and caused changes in the secondary structure of fourth helix.

The secondary structure analysis of ITS2 of *A. hypanicum* and *A. compactum* (fig. 3) demonstrated a total identity of primary and secondary structures of 1st and 2nd helices within a whole sampling. Two ambiguous nucleotides (Y in 40-th position and W in 80-th position) were found for the sample from Turkey (FJ384028).
In the variable part of ITS2 3-rd helix (position №130) the nucleotide substitution was present in two samples of *A. compactum* (FJ384030, Georgia та FJ384032, “Ukrainian cultivar”), but the secondary structure of third helix remained identical for both samples.

Based on the sequence of fourth helix of ITS2 a total sampling can be divided into 3 groups according to the type of nucleotide in position № 189: the sequences of *A. hypanicum* and *A. compactum* from Turkey (FJ384028, FJ384029), Macedonia (FJ384031) and original
sequence of the sample from Russian Federation had T in position № 189, whereas the “Ukrainian cultivar” of \( A. \ compactum \) (FJ384032) had C in the same position, and sequence of the sample from Georgia (FJ384030) contained ambiguous nucleotide (C or T).
A. *hypanicum*. Moreover, according to the above mentioned literature data, *A. compactum* and *A. hypanicum* can be distinguished by the shape of calyx tooth (obtuse tooth for *A. compactum* and acute – for *A. hypanicum*). On the contrary, some herbarium specimens from Caucasus, identified as *A. compactum*, had acute calyx tooth.

To summarize, the population of *A. compactum* from Caucasus can be distinguished from the population of *A. hypanicum* from the territory of Ukraine only by the bacts and apical leaves.

**Discussion**

The absence of any CBC (compensatory base changes) between *A. hypanicum* and *A. compactum* according to A. Coleman concept claims the absence of genetic barriers for breeding between these taxa [COLEMAN, 2003, 2007, 2009].

The analysis of the secondary structures of ITS1 and ITS2 transcripts confirmed three distinct variants of the first and two variants of the fourth ITS1 helices as well as two variants of ITS2 fourth helix (fig. 4). According to these data, the investigated sampling can be divided into 4 main operation taxonomic unites (OTU) (tab. 3).

**Рис. 4. Варіанти вторинних структур першої та четвертої спіралей ITS1, а також четвертої спіралі послідовності ITS2 *Atoction hypanicum* (Klokov) Tzvelev та *A. compactum* (Fisch. ex Hornem.) Tzvelev**

**Fig. 4. A. hypanicum** and *A. compactum* (Fisch. ex Hornem.) Tzvelev secondary structure variants of ITS1 first and fourth helixes and of ITS2 fourth helix

Unique base change in *A. hypanicum* ITS1 fourth helix (site № 210) causes changes in the secondary structure and distinguish it from the other sequences of *A. compactum*. The second OTU is represented by the original sequence of *A. compactum* from Russian Federation and by the sequences from Macedonian (FJ384031) and Turkish (FJ384028) samples. The third OTU is represented by the sample from Turkey (FJ384029), and the fourth one – by “Ukrainian cultivar” (FJ384032). Although, the sample from Georgia (FJ384030) contains Y in position 189 of ITS2 and, therefore, can be placed into the both variants of helix
structure, but with higher probability it belongs to OTU2 than to OTU4 on the basis of differences in first helix of ITS1.

Finally, rating of *A. hypanicum* as the separate OTU and also presence of morphological differences from *A. compactum* provide evidence that *A. hypanicum* is a separate endemic species.

Since *A. compactum* was described from the territory of Russia [FLORA…, 1936], the second OTU most likely represents a classic *A. compactum*. The sample of *A. compactum* (FJ384029), originated from Turkey, is not identical to another Turkish sample (FJ384028) and represents the separate OTU. Possibly, this sample fit *S. vandasii*, which is considered as a synonym of *A. compactum* now. But since we had no specimens from locus classicus of *Silene vandasii* there was no opportunity to investigate them.

**Table 3**

<table>
<thead>
<tr>
<th>ITS1 and ITS2 helices' variants and corresponding operational taxonomic units</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>ITS1-H1</strong></td>
</tr>
<tr>
<td>-----------------</td>
</tr>
<tr>
<td><em>A. hypanicum</em> (UA) KJ616753</td>
</tr>
<tr>
<td><em>A. compactum</em> (RU)</td>
</tr>
<tr>
<td><em>A. compactum</em> FJ384031 (MC)</td>
</tr>
<tr>
<td><em>A. compactum</em> FJ384030 (GE)</td>
</tr>
<tr>
<td><em>A. compactum</em> FJ384028 (TR1)</td>
</tr>
<tr>
<td><em>A. compactum</em> FJ384029 (TR2)</td>
</tr>
<tr>
<td><em>A. compactum</em> FJ384032 (UAc)</td>
</tr>
</tbody>
</table>

“Ukrainian cultivar” (FJ384032) also represents the separate OTU, but the origin of this specimen remains unknown. [FRAJMAN et al., 2009]. Whereas herbarium sample, collected near Sevastopol (KW № 014166) is identical by morphological features to *A. compactum* from Caucasus, we assume that this species is cultivated on the territory of Ukraine. But the origin of the specimen FJ384032 as “Ukrainian cultivar” [FRAJMAN et al., 2009] exactly from Ukraine is still under question.

**Conclusions**

The comparative search of 29 herbarium specimens of *A. compactum* from Caucasus and *A. hypanicum* from the valley of Pivdenny Bug confirmed the presence of morphological differences, connected with structure of bracts and apical (upper) leaves between the two taxa. We determined that features such as tooth shape of calyx, bract length and ratio between calyx and capsule length, proposed earlier for discrimination of these species, cannot be used as diagnostic and discriminative.

Provided reconstruction of ITS1 and ITS2 secondary structures has shown that there were no CBC between *A. hypanicum* and *A. compactum*. This fact supports the absence of any genetic barriers for breeding according to A. Coleman concept.

*A. hypanicum* can be separated from all investigated *A. compactum* sequences by the structural nucleotide substitution in the 4-th helix of ITS2.

Thereby, provided comparative morphological and molecular genetic study confirmed *A. hypanicum* as a separate species and its endemism.

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The authors express gratitude to Sergei L. Mosyakin, Prof., Dr.Sci. Corr. Member, Natl. Acad. Sci. of Ukraine, Director, M.G. Khodolny Institute of Botany and to Nataliia M. Shyyan, PhD, curator of National herbarium of Ukraine (KW) for providing a specimen of *A. hypanicum*.  

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References


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